BCWA PROGRAM GUIDLELINES & OPERATIONS

Draft December 11, 2019



PGO34- BCWA Surface Water Monitoring Program and Sample Analyses Plan Version 2020.01



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Bear Creek Watershed Monitoring Program

Bear Creek Reservoir Control Regulation #74

The Bear Creek Watershed (Figure 1) is a specific geographic area identified in the Bear Creek Watershed State Control Regulation (Regulation #74, 5 CCR 1002-74) (Control Regulation) identifying water quality management (BCWA Policy 13 – Vision, Mission & Targets). The watershed includes all tributary water flows that discharge into Bear Creek Reservoir and a small area below the dam as defined in *BCWA Policy 14 – Watershed Boundary*. Essentially, the watershed boundary extends from the Mount Evans Wilderness on the western end past the Town of Morrison on the eastern end. The two major tributaries are Bear Creek and Turkey Creek. The purpose of the Control Regulation is to attain site-specific water quality standards and classifications through control of total phosphorus and chlorophyll.

The Bear Creek Watershed Association (Association) oversees implementation of the Control Regulation. The Association is the local water quality agency responsible for monitoring and tracking water quality in the Bear Creek Watershed. The Association has conducted a generally continuous collection of surface quality data from 1990 for the Bear Creek Watershed and reservoir. Data collection includes specific chemical, physical and biological parameters. Data is collected monthly and bi-monthly at Bear Creek Reservoir and along Turkey Creek and Bear Creek. The Association meets water quality data sampling and analyses objectives established in the Bear Creek Reservoir Control Regulation # 74.

The Association provides watershed reporting as posted on the Association Website <u>www.bearcreekwatershed.org</u>, which serves to keep federal, state, and local governments and others informed on the state of the watershed. The Control Regulation defines specific reporting requirements, which helps the Association keep the Water Quality Control Commission and Water Quality Control Division staff updated on progress of the Association in implementing the Control Regulation.

Figure 1 Bear Creek Watershed



The Association has established a series of management policies and strategies to guide the watershed monitoring programs (See PGO01 for a complete list of program documents as incorporated into the Association Watershed Plan; and PGO02 for the Program Document Categories).

Monitoring requirements established by State Control Regulation #85 are designed to evaluate the effectiveness of and to determine the sources and load of nutrients at selected locations, and eventual implementation of appropriate and necessary source controls. The Bear Creek Association watershed monitoring plan includes monitoring elements for wastewater treatment facilities in the watershed, which allows these facilities to meet monitoring requirements in Regulation #85.

Nutrient Regulation #85

All wastewater treatment plants in the watershed are categorized as minor facilities. As such, upstream and downstream monitoring is exempt for the treatment works that surface discharge. All surface discharging wastewater treatment facilities must meet the effluent monitoring requirements of Regulation #85. Each facility has certification

and reporting requirements defined in that regulation. The monitoring data collected for Regulation #85 is not part of the permit DMR reporting.

Implementation of Regulation #85 monitoring is a requirement of permitted surface water dischargers and is not a specific monitoring requirement of the Association. However, the Association board has determined that integrating the monitoring requirements from Regulation #85 into the Association watershed monitoring plan Regulation #74 could serve to improve water quality management in the watershed and have a shared funding benefit. A combined monitoring effort can meet state requirements, but, as importantly, it is an opportunity for the Association to develop a more comprehensive and holistic nutrient management plan that will protect and improve water quality throughout the watershed. Involvement of wastewater treatment facilities in this joint monitoring effort is strictly voluntary. See *BCWA Information Sheet 18 Reg 85 Shared Service Program* for shared service program details.

The Association completed the certification process for all wastewater treatment facilities in the watershed and maintains a copy of the certification in the association data record. However, the Association will only do the reporting requirement for those treatment facilities participating in the voluntary shared service monitoring program. All other non-participating treatment plants are expected to do their own reporting to the WQCD in accordance with the Regulation #85 reporting requirements. The wastewater treatment plants that have volunteered for this sampling effort are shown in Table 1. The Association will pick-up collected effluent samples in coordination with these treatment plants. The Association contract laboratory will process these samples. The eight participating treatment plants will compensate the Association for the actual laboratory costs. Data collection will occur on the first or second week of every other month beginning in January.

	Coordinated with Monthly	Not	
Wastewater Treatment Works	Stream Samples	Participant	
Bear Ci	reek Drainage		
JCS Outdoor Lab	X		
Brookforest Inn		х	
Evergreen Metro District	X		
West Jefferson County Metro District	X		
Kittredge Sanitation and Water District	X		
Genesee Water and Sanitation District		X	
Forest Hills Metro District	X		
Morrison	X		
Turkey (Creek Drainage		
Aspen Park Metro District		X	
Conifer Metro District		No Sample	
Conifer Sanitation Association	X		
JCS Conifer High School	X		
Tiny Town		No Sample	
Geneva Glen		No Sample	

Table 1 Wastewater Treatment Plants Involved in a Coordinated Monitoring Plan

Association Monitoring Program Types

The Bear Creek Watershed Association maintains five types of water quality and other monitoring efforts to characterize water and environmental quality within the Bear Creek Watershed:

P1- Routine water quality monitoring at Bear Creek Reservoir (multiple vertical stations), Turkey Creek inflow to reservoir, Bear Creek inflow to reservoir, reservoir discharge into lower Bear Creek, and the lower edge of the watershed near Wadsworth. The P1 sites are long-term reference monitoring sites consistent with the intent of the monitoring program outlined in the Bear Creek Reservoir Control Regulation #74.

P2- Supplemental sampling of tributaries, problem areas, restoration or other project specific sites (e.g., Coyote Gulch in cooperation with the City of Lakewood). These types of monitoring efforts can be either of limited duration, or long-term on a site-specific basis, and generally these programs monitor for specific parameters of interest to the project.

P3- Watershed surface water monitoring along Bear Creek and Turkey Creek drainages for site-specific characterizations (e.g., temperature trends, nutrient loading, flow studies). These are interim and long-term monitoring sites for watershed characterizations. Watershed monitoring stations include both long-term reference sites where multi-year data is desirable, and target sites that may provide only a couple years of data. The nutrient monitoring is on a watershed basis that begins near Summit Lake and extents through Bear Creek Reservoir. The monitoring schedule for the watershed programs are maintained by the Association manager and are available on request.

P4- Supplemental environmental characterizations of Bear Creek watershed including, but not limited to macroinvertebrates, flow analysis, habitat characterizations, fishery evaluations, system productivity, or other environmental factors that potentially affect fisheries or watershed health.

P5- Wastewater treatment facility nutrient sampling consistent with Regulation #85. The wastewater treatment plants in the watershed are listed in Table 2.

Wastewater Treatment Plants	Facility	Design	2020 Effluent
	Туре	Capacity MGD	Monitoring
JCS Outdoor Lab	Minor	0.0075	Yes
Brookforest Inn	Minor	0.009	No
Evergreen Metro District	Minor	0.99	Yes
West Jefferson County Metro District	Minor	0.7	Yes
Kittredge Sanitation and Water District	Minor	0.125	Yes
Genesee Water and Sanitation District	Minor	0.8	Yes
Forest Hills Metro District	Minor	0.05	Yes
Morrison	Minor	0.35	Yes
Aspen Park Metro District	Minor	0.025	Yes
Conifer Metro District	Minor	0.043	Yes
Conifer Sanitation Association	Outfall	0.019	Yes, CMD
JCS Conifer High School	Minor	0.052	Yes
Tiny Town	Minor	0.005	No
Geneva Glen	Minor	0.0105	Yes

 Table 2
 WWTF in Bear Creek Watershed

Mapped Watershed Features

The BCWA uses Google Earth to track monitoring sites and watershed features of interest. The mapping codes and station types are shown in Table 3. A Google Earth *Myplaces* file is available to the membership for use with Google Earth.

Table 3	BCWA Google Earth Station Types
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Code		Station Type	
P1	P1 Routine CR Monitoring stations		
	P1 BCR	BCR Stations	
	P1 S	Stream Sites for mass balance	
P2		Supplemental Water Quality Characterizations in Bear Creek Watershed	
	P2 Air	BCR Aeration Sites	
	P2 Coyote	Coyote Gulch Nutrient Reduction	
	P2 Fen	Summit Fen Study	
	P2 Gen	Genesee Reservoir	
P2 K/S Kerr and Swede Gulch Special Study		Kerr and Swede Gulch Special Study	
P2 MV Mount Vernon Creek		Mount Vernon Creek	
P2 Sed Bear Creek Reservoir Sediment Study		Bear Creek Reservoir Sediment Study	
P2 Soda Soda Lakes		Soda Lakes	
	P2 Summit	Summit Plume Study	
	P2 Temp	Stream Temperature Only Probe	
	P2 Trib	Special tributary Nutrient loads	
	P2 Trouble	Troublesome Gulch	
	P2 TSS	Total Suspended Sediment Sites	
	P2 Flow	Stream Flow Measurement Sites	
	P2 Copper	Segment 1e Copper Study	

P3	Watershed Monitoring Program Active sites		
P3 EGL	Evergreen lake		
P3 WS	Watershed		
P3 Old	Stream Site Not Active		
P4	Supplemental Studies, Education Sites or Informational Features		
P4 Ed	Educational site		
P4 Fish	CDFW Fishery Surveys		
P4 Gage	Gaging Stations		
P4 Horse	P4 Horse Horse Stables or Operations		
P4 Macro	Macroinvertebrates		
P4 Other Group	Other Outside Agency Monitoring Stations		
P4 Sign	BCWA Signs		
P4 Segments	Mainstem segment brake points		
P4 WS	Watershed Features		
P4 Weather	Watershed Weather Stations		
P5	WWTF		
P5 Outfall	WWTF Outfalls		

2020 Water Quality Monitoring Program & Quality Assurance Project Plan

The following monitoring plan sections details the 2020 reservoir and watershed monitoring programs as approved by the BCWA Board. This monitoring plan remains consistent with the quality assurance goals of the previously adopted Association QAPP (Bear Creek Watershed Association, 2006). However, this monitoring plan is the working version. The monitoring program version 2020.01 is adapted from the last version of the 2019.02 monitoring plan.

Watershed Field Monitoring Personnel

The Association contracts field monitoring, sampling coordination, and data management with RNC Consulting LLC:

Russell N Clayshulte, RNC Consulting LLC 1529 S Telluride St Aurora, CO 80017 Office (303) 751-7144 / Cell (303) 638-4931 rclayshulte@earthlink.net

RNC Consulting LLC conducts field sampling in cooperation with Evergreen Metropolitan District (EMD) and the City of Lakewood, and occasionally other members. EMD staff assist with field monitoring, temperature probe maintenance and the annual data report. While several staff of EMD assist with the monitoring program, the EMD primary field manager/ data manager is:

Tony Langowski, Evergreen Metro District Office (303)674-4112 Cell (303) 301-5507 tlangowski@evergreenmetrodistrict.com

The City of Lakewood assists with sampling on Coyote Gulch, Bear Creek Reservoir, and P1 sites. While several staff of Lakewood Bear Creek Lake Park staff assist with the monitoring program, the Lakewood primary field monitoring assistant is:

Mike Towner, Bear Creek Lake Regional Park Cell (303) 941-7554 <u>miktow@lakewood.org</u>

All water quality samples for laboratory analyses are delivered by RNC Consulting LLC to GEI Consultants, Inc. / Chadwick Ecological Division within 2 hours of final sample collection.

Field data sheets are scanned and converted into PDF files for electronic storage. Field data is transferred into a master spreadsheet, which contains all annualized data collected for the Association, except for temperature logger data.

BCWA Field Methods

The BCWA has adopted a set of indices forms, field methods and habitat summaries, which are listed and maintained in the PGO1 Master Index List as part of the Association's Watershed Plan (Table 4).

Table 4 BCWA Indices & Methods

	BCWA Indices & Methods
FI01	BCWA Habitat Indices Form
FI02	BCWA Physical Stream Indices Form
ME01	Embeddedness Field Estimation Method
ME02	Gravelometer Pebble Count Method
ME03	Field Flow Estimation Method
ME04	Macroinvertebrate Field Sample Method
ME05	Periphyton Field Estimation Method
ME06	Water Clarity Estimation Method
ME07	Habitat Indices Site BCW
ME08	Erosion Pin Method (Not Currently Used)
ME09	2015 Habitat Indices Summary
ME10	Tributary Assessment Procedure
ME11	Temperature Probes
ME12	Macro Labels

Watershed Field Monitoring Notes and Methods

Treatment facilities listed in Table 2 generally provide effluent data collected at their respective WWTP's, including analytical results; and /or assistance with special monitoring.

The Interval frequency for temperature dataloggers is every half-hour (48 per day). The dataloggers in wastewater plant discharges will be field-downloaded as needed, 60-90 days, while dataloggers located in the stream will be field-downloaded as needed.

Manual pH, Temperature, DO, Specific Conductivity, Total Nitrogen, Nitrate-nitrogen, Total Ammonia, Total Inorganic Nitrogen and Total Phosphorus data collected at selected temperature datalogger locations. Manual monitoring is performed in the morning to early afternoon, beginning at approximately 08:00 and ending at approximately 15:00. Monthly monitoring provides a check on integrity of dataloggers.

WWTP effluent data collected as part of the typical plant process control performed daily. Effluent pH/Temperature/DO recorded and ammonia sampling performed in accordance with the requirements of each WWTP discharge permit. Total ammonia analyzed for WWTP effluents by the method allowing for the lowest detection limit.

USGS flow measurement obtained at gages above Evergreen Lake, above the Town of Morrison and on Turkey Creek.

Daily weather data (High/low temperature, precipitation) from the NWS station at the EMD WWTP obtained on a monthly basis.

Calibrations of portable equipment documented prior to each use and Certificates of Calibration for all equipment obtained. NIST certifications and Certifications of Compliance are originally obtained for each temperature datalogger used in the study.

GPS points maintained for all new sampling and monitoring locations.

RNC Consulting LLC works with the City of Lakewood to monitor dissolved oxygen in water column in Bear Creek Reservoir and adjust the operation of the reservoir aeration system to maintain DO standards, while minimizing aeration operations. This may require additional vertical probe sampling in the July to October period to monitor DO levels in the water column at site 40.

RNC Consulting LLC maintains photographic points for critical segments and conditions, and documents dewatering of Bear Creek Segment 1b below both the Arnett-Harriman and Ward ditches.

All collected data is maintained in an annual master spreadsheet, which is posted at the completion of the annual monitoring program on the Association Website.

Contract Laboratory and Laboratory Methods

The contract laboratory used by the Association for all laboratory analysis is GEI Consultants, Inc. / Chadwick Ecological Division.

Natalie Love, Laboratory Manager GEI Consultants, Inc. Ecological Division 4601 DTC Boulevard, Suite 900 Denver, CO 80237

Macroinvertebrate Analysis

GEI Consultants analyses samples by the BCWA for benthic macroinvertebrates. Samples collected by the Association follow the *BCWA M04 Macroinvertebrate Field Sample Method*. Data for samples are reported as number of organisms per square meter. Percent of total is also reported. The species are counted consistent with the CDPHE EDAS import columns for taxa and species. This data is converted into MMI scores using the EDAS protocols (EDAS CO Master Version - Distributable to 3rd Parties and *A Stepwise Guide to Generating MMI Scores Using Colorado-EDAS*). Table 5 shows the Macroinvertebrate QA performance from the GEI Laboratory.

Table 5	Macroinvertebrate	QA	Performance
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	Exti	ractions	Initial Id	entifications
# of Samples for	Sample %	Acceptable %	Sample %	Acceptable %
1	98.1	95	99.3	95

Laboratory Methods

The GEI Laboratory methods and limits are summarized as follows:

- Analyte Methods are shown in Table 6.
- General Preservation: preserved with H2SO4 to pH <2.0
- Storage: refrigerated at 4°C until analyzed
- Number of Replicates/Analyte: 2 for all nutrients
- QA/QC Analyses: 3 standards analyzed at beginning and end of analyses, and at 5 replicate increments, for all nutrients
- Example of a Matrix Spike by analyte report is shown in Table 7.
- Table 8 reports the minimum detection limits used by GEI.
- Table 9 shows the New GEI Lachat Method for Nutrients.
- Nutrient Analysis
 - \circ Preservation: preserved with H₂SO₄ to pH <2.0
 - Storage: refrigerated at 4°C until analyzed
 - Number of Replicates/Analyte: 2 for all nutrients
 - QA/QC Analyses: 3 standards analyzed at beginning and end of analyses, and at 5 replicate Increments, for all nutrients
- Total Suspended Solids Analysis
 - Storage: refrigerated at 4°C until filtered
 - ο Filtration: filtered through a pre-baked Gelman A/E 1.0 μm glass fiber filter
- Chlorophyll a Analysis
 - ο Filtration: filtered through a pre-baked Gelman A/E 1.0 μm glass fiber filter
 - Storage: filter stored frozen at -20°C, kept in the dark and in desiccator
- E. coli Analysis
 - All testing would follow the Hach colilert method for E. coli adapted from Standard Methods (9223 B) for the Examination of Water and Wastewater (APHA et al, 1998).
 - Sample containers supplied by GEI.
- Copper Study
 - EPA 200.8, filtered and digested samples
 - Hardness Standard Methods 2340 C

Table 6 **Analyte Methods**

Analyte	Method	Filtered (0.45 µm filter)	Preservation (H2SO4 to pH <2.0)	Allowable Hold Time (unpreserved samples) *
Total Phosphorus	QuickChem 10-115-01-4-B, with manual digestion	No	Yes	48 hours prior to digestion; 7 days prior to analysis
Total Dissolved Phosphorus	QuickChem 10-115-01-4-B, with manual digestion	Yes	Yes	48 hours prior to digestion; 7 days prior to analysis
Total Nitrogen	QuickChem 10-107-04-4-B, with manual digestion	No	Yes	48 hours prior to digestion; 7 days prior to analysis
Nitrate + Nitrite	QuickChem 10-107-04-1-C	Yes	No	48 hours
Ammonia	QuickChem 10-107-06-2-A	Yes	Yes	24 hours
Total Suspended Solids	Standard Methods 2540 D	Yes		7 days before filtration, indefinitely after drying
Chlorophyll a	Hot Ethanol Extraction	Yes		28 days
E. coli	Hach colilert method (9223 B)	No	No	24 hours
Copper	EPA 200.8	Yes	No	48 hours
Hardness	Standard methods 2340 C	No	No	48 hours

*all preserved samples have an allowable holding time of 28 days

Example of Matrix Spikes by Analyte Reported to Association. Table 7

Analyte	Average % Recovery	Number of Samples Spiked	Within Target Range			
TP	106	1	Yes			
TDP	103	1	Yes			
TN	102	1	Yes			
NOx	98	1	Yes			
NH3	93	1	Yes			
Target rec	Target recovery range is 80-120% recovery.					

Table 8 **Minimum Detection Limits**

Analyte	Method	MDL (ug/l)	PQL (ug/l)
Total Phosphorus	QuickChem 10-115-01-4-B, with manual digestion	2 ug/l	8
Total Dissolved Phosphorus	QuickChem 10-115-01-4-B, with manual digestion	2 ug/l	8
Total Nitrogen	QuickChem 10-107-04-4-B, with manual digestion	6 ug/l	42
Nitrate+Nitrite	QuickChem 10-107-04-1-C	2 ug/l	8
Total Ammonia	QuickChem 10-107-06-3-A	5 ug/l	35
Total Suspended Solids	Standard Methods 2540 D	4 mg/l	
Chlorophyll a	Hot Ethanol Extraction	0.1 ug/l	
Copper	EPA 200.8	0.26 ug/l	
Hardness	Standard Methods 2340 C	2 mg/l	

The GEI Consultants, Inc. (GEI) Laboratory uses a Lachat instrument and accompanying methods used for low-level nutrient analyses. All nutrient samples are analyzed using GEI's Lachat methods (Table 8). All methods are either EPA-accepted, EPA-equivalent or have been approved by the state of Colorado for use in GEI lab. All methods are acceptable under Regulation 85 (Nutrients Management Control Regulation). GEI will continue to follow the same rigorous QA/QC procedures they have always followed under our previous methodology.

Analyte	Old GEI Method	New GEI Method	MDL ug/l	PQL ug/l
NH3	QC 10-107-06-3-D	QC 10-107-06-2-A	5	35
NOx	QC 10-107-04-1-B	QC 10-107-04-1-B	2	14
NO2	QC 10-107-04-1-B	QC 10-107-04-1-B	2	14
TN/TDN	SM 4500-N B (mod)	QC 10-107-04-4-B	6	42
OP	QC 10-115-01-1-T	QC 10-115-01-1-T	2	14
TP/TDP	OC 10-115-01-4-U	OC 10-115-01-4-B	2	14

Table 9 Lachat Method for Nutrients

Laboratory QA/QC Protocols for Nutrient Analyses

Equipment calibrations performed each time new standards are prepared (minimum of once per week). If the r-value of the standard curve is less than 0.999, the instrument is recalibrated, or standards are remade. Replicates run on each sample are analyzed and the percent difference must be within 10% if the resultant concentration is above the

minimum detection limit. If results of analyses of replicate samples are not within 10% of one another, samples are placed in a clean test tube and reanalyzed.

During analysis, check standards are analyzed between every 5 samples (or 10 replicates). The check standards consist of one high range standard, one mid-range standard, and the zero (blank). Check standards analyzed before and after each group of samples must be within 10% of the theoretical value. If standards are outside of this range, samples and standards are placed in clean test tubes and reanalyzed to try to determine the source of the problem. Sample values are not accepted until the problem has been resolved and all check standards pass the QC criteria. One matrix spike is run for every 10 samples analyzed. The percent recovery for matrix spikes must be $\pm 20\%$.

After sample analyses a final QC check performed to determine if all parameters measured agree. Final analyses for each sample are compared to ensure that concentrations of total phosphorus \geq total dissolved phosphorus \geq orthophosphate and that the concentration of total nitrogen \geq total dissolved nitrogen \geq nitrate/nitrite and ammonia. If parameters do not agree, samples are reanalyzed.

<u>Chain of Custody</u>

The Association provides the laboratory with a chain-of-custody form with the transfer of samples that identifies each sample, parameters required for sample, date and time of collection, sample personnel and where data is reported. At transfer a staff member from the laboratory signs and dates the chain-of-custody and makes a copy for RNC Consulting LLC. The laboratory uses Table 10 as part of their chain-of-custody.

QA Requirement	Yes	No	Note
Chain of Custody received complete			
Samples received within holding times			
Samples at correct temperature (1-8°C)			
QA/QC Standards within acceptable 10% difference			
Duplicate samples within acceptable 10% difference			
All matrix spikes within target range of 80-120% recovery			

Table 10 Laboratory Chain-of-Custody

Data Management

RNC Consulting LLC is responsible for all Association water quality and environmental data management, including QA/QC of data. Large quantities of varied data are collected during the annual monitoring program: Monthly stream monitoring and sampling, laboratory results, thirty-minute temperature measurements from dataloggers, wastewater treatment plant effluent process, control, and permit monitoring data (from five treatment plants), weather statistics and stream flows comprise raw data. All data are stored on an office computer, using Microsoft Office 365 software. Most of the data resides in and analyses occurred in Excel spreadsheet format. Data record is preserved in an off-site cloud storage system.

The Association data is located at two different locations. Watershed data collected with the assistance of EMD staff is maintained on computer systems at the EMD offices. All raw watershed data electronically forwarded from EMD staff to the Manager for data summary and analyses. RNC Consulting LLC maintains all monitoring data for all Association monitoring programs. Data is kept on a computer with back-up to an external cloud-drive. Additionally, some back-up data sets of recent data are kept on flash drives.

The Association incorporates data into a data report, after the study. Electronic data files are transmitted to the Water Quality Control Division, Colorado Division of Wildlife and Evergreen Trout Unlimited after the Association Board has approved the study report. Sampling and Monitoring Plan summaries are provided to the BCWA monthly at meetings, which are open to the public.

Watershed Sampling Dates 2020

The 2020 monitoring schedule, which maybe periodically updated or changed as required by the field monitoring personnel, is maintained by the Association Manager. Changes to the monitoring schedule are sent to the Association through electronic notifications.

Water Quality Monitoring Parameters in 2020

Table 11 lists the 2020 water quality monitoring parameters by monitoring program.

Watershed and Special Stream WQ Studies					
Field Chemistry/ Physical	Laboratory Analyses				
Temperature (discrete field probe)	Total Nitrogen (GEI)				
Temperature (continuous data loggers, 1/2-2m)	Total Phosphorus (GEI)				
Dissolved Oxygen, YSI Probe	E. coli, sites 16c, 45 and 90 (GEI)				
Specific Conductivity, YSI Probe	Total Suspended Sediments, if needed Spring Runoff				
pH, YSI Probe					
Manual Flow/ gage readings					
Water Clarity - Staining					
Periphyton Coverage					
	ilities Regulation 85 Requirements				
Field Data	Laboratory Analyses (GEI)				
Daily average effluent discharge	Total Nitrogen				
Temperature (continuous data loggers, Effluent)	Nitrate/Nitrite as N, dissolved				
	Total Ammonia				
	Total Inorganic Nitrogen (Calculation = NO2+NO3+NH4)				
	Total Phosphorus				
Monitoring Site Su	rvey - Annual Select Sites				
Macroinvertebrates - 10 sites	Habitat Indices				
Pebble Counts	Physical Stream Indices				
Embeddedness					
BCR and EGL Sediment	t Survey - Annual Limited Sites				
Sediment TP (Pore Water): BCR 6 sites, EGL 1 site	% Total Solids				
% TOC (Organics)					
Reservoirs (E	BCR and Evergreen)				
Field Data	Laboratory Analyses				
Temperature (field probe, 1/2-m intervals in central pool)	Total Nitrogen (-1/2m and +1m) (GEI)				
Temperature (continuous data loggers, 1/2-2m)	Total Phosphorus (-1/2m and +1m) (GEI)				
Dissolved Oxygen (field probe, 1/2-m intervals in central	Chlorenhull o (1/2m only) (CEI)				
pool thru 4m, then 1m interval)	Chlorophyll a (-1/2m only) (GEI)				
Specific Conductivity (field probe, 1/2-m intervals in	BCR Phytoplankton (June, July, August, September, October)				
central pool thru 4m, then 1m interval)	EGL (July, August, September) (GEI)				
pH (field probe, 1/2-m intervals in central pool thru 4m,	Zooplankton - annual, species present (GEI), if needed				
then 1m interval)	Zoopiankion - annual, species present (OE1), it needed				
Total Depth					
Secchi Reading					

2020 Monitoring Stations and Frequency of Sampling

Table 12 lists the 2020 monitoring stations, type of monitoring, reference sites and frequency of sampling by stream segments.

Table 122020 Monitoring Stations

	Watershed Sites by Stream Segment	2020						Site
Site ID		Temp Logger	Flows	Field Probe	TN/TP	Fishery	Macroinvertebrates	Туре
	Segment 1a							
Site 58	Bear Creek Below Mt. Evan Wilderness	х	х	х	х		Х	P3 WS
Site 2a	Golden Willow Bridge	х	х	х	х	х	Х	P3 WS
Site 3a	te 3a Above Evergreen Lake at CDOW Site		USGS	х	х	x	Х	P3 WS
Segment 1b								
Site 15a	Bear Creek within Bear Creek Park	х	USGS	х	х		Х	P1 BCR

					2020	1		Site
Site ID	Watershed Sites by Stream Segment	Temp Logger	Flows	Field Probe	TN/TP	Fishery	Macroinvertebrates	Туре
WWTF Site 24	Morrison WWTF	х			R85			P5
		Segmer	nt 1c	1	T	1	1	P
Sites 40, 41, 42	Bear Creek Reservoir	X		X	х			P1 BCR
	T	Segmen	t 1d		1	1	ſ	
Site 4a/4e	Evergreen Lake	X		X	х			P3 EGL
	T	Segmen	t 1e		1	1	ſ	
Site 5a	Evergreen Little Bear site at CDOW	X	x	X	x	x	х	P3 WS
Site 9	O'Fallon Park, west end at CDOW Site	X	x	X	x	x	х	P3 WS
Site 12	Lair o' the Bear Park, at CDOW site	X	x	X	x	x	х	P3 WS
Site 13a	Below Idledale, Shady Lane CDOW site	X	x	X	x	x	х	P3 WS
Site 14a	Morrison Park east end at gaging station	X	USGS	X	x	x	х	P3 WS
WWTF Site 20	Evergreen Metro District	X			R85			P5
WWTF Site 22	Kittridge Water & Sanitation	X			R85			P5
WWTF Site 23	Genesee Water & Sanitation District	x						P5
	1	Segme	nt 2		1	1	ſ	
Site 45	Lower Bear Creek, below reservoir trace	X	DNR	X	x			P1 BCR
Site 90	Lower Bear Creek Wadsworth	X	USGS	X	х		Х	P3 WS
		Segme	nt 3	1	1	1	1	1
Site 25	Vance Creek	х	х	X	х			P3 WS
WWTF Site 71	JCS Outdoor Lab	х			R85			P5
		Segmen	it 4a		T	1	1	r
Site 47a	Upper Coyote Gulch		х	х	x			P2 Coyote
Site 47b	Lower Coyote Gulch, Above Reservoir		x	x	x			P2 Coyote
		Segmer	nt 4c			1		
WWTF Site 34a	Forest Hills Metro District	x			R85			P5
Site 34b	Mt Vernon Drainage, Mathew-Winter	х	х	x	х		Х	P3 WS
		Segme	nt 5	•				
Site 26	Cub Creek, at mouth	х	х	х	х			P3 WS
Site 32	Troublesome at mouth		x	x	х			P2 Trouble
Site 64	Troublesome at Culvert above West Jeff		x	x	x			P2 Trouble
WWTF Site 21	West Jefferson County Metro District	х			R85			P5
		Segmer	nt 6a		T	1	1	r
Site 16a	Turkey Creek within Bear Creek Park	х	х	Х	х			P3 WS
Site 18	South Turkey Creek Myers Ranch	х	х	X	х			P3 WS
		Segmen	t 6b					
WWTF Site 73	Conifer Sanitation District				R85			P5
WWTF Site 59	JCS Conifer High School				R85			P5
Site 19	North Turkey Creek Flying J Ranch	х	х	х	х			P3 WS
	5	Segments '	7 and 8			1		r
Site 36	Summit Lake (Segment 8)		х	х	х			P3 WS
Site 37b	Bear Creek Mainstem (Segment 7)	х	х	x	x			P3 WS
Site 63	Special Fen Study			x	х		P2 Fen	
	Segments 10, 11	l, and 12 (Lake/ Po	nd Segn	nents)			
	None in 2019							

Wastewater Treatment Facility Regulation #85 Monitoring

The Bear Creek Association watershed monitoring plan includes monitoring elements for wastewater treatment facilities in the watershed, which allows these facilities to meet monitoring requirements in Regulation #85. The monitoring data collected for Regulation #85 is not part of the permit DMR reporting.

The wastewater treatment effluent parameters used to meet Regulation #85 requirements are shown in Table 13. The PQLs and MDLs are lower than those required in Regulation #85 and are shown in Table 14. These MDLs and PQLs are used for all Association nutrient data processed by the Associations contract laboratory.

Wastewater Treatment Facilities				
Field Data	Laboratory Analyses			
Daily average effluent discharge	Total Nitrogen			
Temperature (Selected plants continuous data loggers, Effluent)	Nitrate+Nitrite-Nitrogen			
	Ammonia-Nitrogen			
	Total Inorganic Nitrogen (Calculation = NO2+NO3+NH4)			
	Total Phosphorus			

Table 13Regulation 85 Parameters

Table 14MDLs and PQLs used for Association Sample Parameters

			MDL	PQL
Analyte	Old GEI Method	GEI Method	(ug/l)	(ug/l)
NH3	QC 10-107-06-3-D	QC 10-107-06-2-A	5	35
NOx	QC 10-107-04-1-B	QC 10-107-04-1-B	2	14
NO2	QC 10-107-04-1-B	QC 10-107-04-1-B	2	14
TN/TDN	SM 4500-N B (mod)	QC 10-107-04-4-B	6	42
OP	QC 10-115-01-1-T	QC 10-115-01-1-T	2	14
TP/TDP	QC 10-115-01-4-U	QC 10-115-01-4-B	2	14

The monitoring frequency is to have 6-monthly samples for treatment plants per year. The State is interested in winter numbers and evenly spaced effluent data. The sample months for Regulation #85 sampling are January, March, May, July, September, November. Data collection will occur on the first or second week of every other month beginning in January 2020.

The Association stream flow monitoring program and analysis plan is enough to meet the intent of Regulation #85 and allow the Association to mass-balance nutrients in the watershed

Small treatment plant effluent samples can be grab samples, if defined as such in permit. If a larger plant is required by permit to do composites, then the sample used for Regulation #85 should be a composite that matches permit requirements. Sampling for nutrients is required in the effluent before it is discharged into the receiving water body at the location where monitoring is performed to satisfy other CDPS permit requirements (as per regulation). Total phosphorus data collected under Regulation #85 for small treatment plants may be used to meet Regulation #74 requirements. The nutrient data collected under Regulation #85 are not required to be reported by the permittee in their respective NPDES DMR reporting system; however, the collected data can be submitted as part of the DMRs. Each plant is responsible for getting the daily average effluent discharge and reporting this information to the Association on a monthly basis.

The Association will provide the necessary sample bottles to the treatment plants for sample collection that are participating in the cost share program. The treatment plant operators must collect necessary effluent samples. The Association cannot take the samples at the plants.

Treatment Tech, who operates smaller treatment works, is opting out of the Association shared monitoring program. The Association will not be responsible for the annual submittal and data transfer to the state for those treatment facilities not participating in a joint monitoring program.

A certification letter is available for each treatment plant. The Association bundled the available certifications, noting the facility is covered by a watershed monitoring program. The Association maintains a copy of the plant certifications to link with the monitoring plan. The nutrient data collected under the regulation #85 regulations are not required to be reported by the permittee in their respective NPDES DMR reporting system; however, the collected data can be submitted as part of the DMRs. See Table 15 for certification status.

Wastewater Treatment	Permit	LRP Certification	Effluent Sample	Copy of	Latitude/Longitude
Plants	Number		Туре	Certification	
		Bear Cree	k Drainage		
JCS Outdoor Lab	CO-0032514	Kim Brogan	grab	Yes 2016	To be determined
Brookforest Inn	CO-0030261	Robert Clodfelter	grab	Yes	39.579394/ 105.380764
Evergreen Metro District	CO-0031429	Dave Lighthart	Composite12-hour	Yes	39.38' 16.19/105.18' 56.07
West Jefferson County Metro District	CO-0020915	Dave Lighthart	Composite12-hour	Yes	39.39' 46.05/105.20' 06.62
Kittredge Sanitation And water District	CO-0023841	Dave Lighthart	Composite12-hour	Yes	39.39' 27.75/105.17' 15.04
Genesee Water and Sanitation District	CO-0022951	Chris Brownell	Composite 24-hour	Yes	39.40' 34/ 105.16' 26
Forest Hills Metro District	CO-0037044	Bruce McCreary	Composite12-hour	Yes	39.42' 09/ 105.15' 07
Morrison	CO-0041432	John McEncroe	Composite12-hour	Yes	39.39' 10.89/ 10510' 39.99
		Turkey Cre	ek Drainage		
Aspen Park Metro District	CO-0631016	Robert Clodfelter	Composite12-hour	Yes	39.32' 38/ 105.17' 25
Conifer Metro District	CO-047295	Bryan McCarty	Groundwater	Yes	Not Sampled
Conifer Sanitation Association	COX-0047392	Becky Hammer	Grab	Yes	39.31' 49/ 105.18' 16
JCS Conifer High School	CO-047988	Kim Brogan	Composite12-hour	Yes	95.523470/ 105.306350
Tiny Town	CO-0036129	Robert Clodfelter	grab	Yes	39.36' 22/105.13' 38
Geneva Glen	CO-0044652	Ken Atchison	Groundwater	Not Needed	Not Sampled

Table 15	Certifications Available for WWTF under Regulation #85
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P1 - Routine Monitoring Program

The routine monitoring program (P1) focuses on Turkey Creek drainage and Bear Creek drainage inputs and discharge from Bear Creek Reservoir (Figure 2) into lower Bear Creek with a central pool characterization of the reservoir near the dam (Site 40). In Figure 3, the outlet structure is near site 41 with Bear Creek inflow near site 44 and Turkey Creek inflow near site 43. The reservoir chemistry and biological characterization occur at site 40. Vertical probe samples at ½ and 1-meter intervals measured at sites 40, 41, and 42 beginning at -1/2-m. Temperature Logger profile of Bear Creek Reservoir at Site 40 with buoy placement and probes attached at ice-off (April-December, first week): ½ m, 1m, 1 ½m, and 2m. Field probe measurements year-round at site 40 with profile interval of ½ m, 1m, 1 ½m, 2m, 2 ½m, 3m, 3 ½ m, 4m, 5m, 6m, 7m, 8m, 9m, 10m, and 11m. Similar profile pattern used at other reservoir sites. The current monitoring program optimizes data generation to evaluate reservoir inflow loading, tropic state changes within the reservoir, and reservoir outflow; while minimizing monitoring cost.

Field Sampling Management: Russell Clayshulte, Association Manager; Field Assistance from Lakewood, Mike Towner.

2020 Routine P1 sampling Sites

The six 2020 P1 routine watershed-monitoring stations, including the reservoir station, are:

- 1. Mainstem of Turkey Creek prior to discharge into Bear Creek Reservoir, within Bear Creek Park, adjacent to the City of Lakewood Maintenance Yard and upstream of the Denver Water diversion;
- 2. Mainstem of Bear Creek prior to discharge into Bear Creek Reservoir, within Bear Creek Park, adjacent to the bridge at the western edge of the park;
- 3. Tail-water discharge from Bear Creek Reservoir in the concrete channel that starts the lower Bear Creek;

- 4. Bear Creek Reservoir, center of main pool and supplemental vertical profile stations 40, 41, and 42.
- 5. Bear Creek Site 90 above Wadsworth.



Figure 2

2020 P1 Bear Creek Reservoir Monitoring Sites



Figure 3 P1 Reservoir Monitoring Stations; Site 40 is the Nutrient P1 station

P1 Sampling Parameters BCR

Table 16 Bear Creek Reservoir Sampling Parameters

Parameter (units)	Reservoir Sites	Reservoir Outflow, Site 45		
Physical/Field				
Flow/ Discharge (cu m/s)	Manual and Staff gage	Manual and Staff gage		
Specific Conductance (umhos/cm)	(Profiles at sites 40, 41, 42)	Х		
Secchi (meters)	Sites 40, 41, 42			
Total Depth (m)	Sites 40, 41, 42			
Dissolved Oxygen (mg/l)	(Profiles at sites 40, 41, 42)	Х		
	(Profiles at sites 40, 41, 42)			
Temperature (C)	Data Logger at site 40	Х		
Total Suspended Sediments (mg/l)	Seasonal Only Spring	Х		
pH (standard unit)	(Profiles at sites 40, 41, 42)	Х		
Biological (Site 40 only)				
Chlorophyll a (ug/l)	X (-1m)			
Zooplankton (August)	Vertical Tow			
Phytoplankton (June, July, August, September, October)	Composite top 1-meter water			
Nutrients (Reservoir Site 40 only)				
Total Nitrogen (ug/l)	X (top, lower)	Х		
Total Phosphorus (ug/l)	X (top, lower)	Х		
Bottom Sediments BCR				
Total Phosphorus (mg/kg)	6 sites			
% Organics (TOC)	6 sites			

Watershed and Special Stream WQ Studies			
Field Chemistry/ Physical	Laboratory Analyses		
Temperature (discrete field probe)	Total Nitrogen		
Temperature (continuous data loggers, 1/2-2m)	Total Phosphorus		
Dissolved Oxygen, YSI Probe	E. coli, select sites		
Specific Conductivity, YSI Probe	Total Suspended Sediments, select sites Spring Runoff		
pH, YSI Probe			
Manual Flow/ gage readings			
Water Clarity - Staining			
Periphyton Coverage			
Monitoring Site Survey - Annual Select Sites			
Macroinvertebrates	Habitat Indices		
Pebble Counts	Physical Stream Indices		
Embeddedness			

Table 17 Monitoring Parameters for Sites 15a, 16a, 16c, 45 and 90

P1 Sampling Frequency

P1 sites sampled monthly in January, February, March, April, May, June, October, November, and December. Growing season samples taken twice in July, August, and September.

P2 - Supplemental Water Quality Characterizations in Bear Creek Watershed

Addressing WQ Concerns on Tributary Drainages

- 1. 2020 special assessment of potential tributary load areas, as directed by Board.
 - a. Conduct ground surveys to identify potential "hot" spots; map
 - b. Conduct multiple field probe measurements at intervals to see if any discernible filed data is event. Only collect TP/TN pair if a suspected area is found, will need linked flow data to calculate nutrient loads. After spring rainy period.

Coyote Gulch

The Association coordinates with the City of Lakewood a sampling program on Coyote Gulch in the Bear Creek Park. The monitoring is done at two sampling sites: above the restoration project, and at the discharge into the reservoir (Figure 4). The Association collects the chemistry data for total phosphorus and nitrate-nitrogen. The Association takes monthly flow measurements to determine nutrient loading. The Association also collects data for temperature, pH, specific conductance and Dissolved Oxygen. Data results are incorporated into the Association monthly and annual data summaries. The Association has pre-construction and post-construction loading data. This monitoring project has established a total phosphorus trade credit for use of the Association.



Figure 4 Coyote Gulch Sampling Sites

Bear Creek Reservoir Sediment Study

Survey Duration: Annually

Obtained core samples at multiple sites after shut-down of aeration system.

Survey Types: Stratified and Single stage

Discrete dredge samples from each of three reservoir zones; Central pool, Turkey Creek inlet and Bear Creek inlet using three fixed transects (Figure 6). Bottom samples obtained with a petite Ponar sampler. This sampler takes a grab of the top 5-6 cm of the mud bottom. One dredge drop made at each site resulting in about 0.5 liters of bottom mud. The locations in Figure 6 are estimates. Sub-samples are bagged from the composite mud sampled at the site.

Total Organic Matter

About 5 grams of sample of the air-dried sediment was weighed, dried for 1 hour at 103-105 degrees C, reweighed. This value is the total solids content of the sample. The dried sample was then ashed in a muffle furnace at 550 degrees C for 15 minutes and reweighed. This procedure provides a total solids percentage and a total organic carbon or Volatile solid percentage. The total organic matter or total organic carbon (TOC) is volatilized and the percent difference is calculated to determine the estimated percentage of TOC contained in surface bottom muds.

Total Phosphorus

Extraction Procedure - Distilled Water

Weigh 4 g of wet mud into 125 ml bottle with lid, add 20 mL of distilled water and shake manually.

Centrifuge at 3,500 rpm for 15 minutes and filter the solution through a 0.7 µm membrane filter.

Use 5 ml for analysis; retain remainder for dilution, as appropriate. Determine total phosphorus as water extractable P in wet mud using HACH method TNT 843.



Figure 5 Sediment sampling sites

Summit Lake and Plume

Bear Creek Watershed Association established two sampling stations at Summit Lake and upper Bear Creek, Mt Evans Wilderness, Clear Creek County Colorado. The Association selected sampling Site 36 (Summit Lake at outfall) and Upper Bear Creek Site 37 to monitor assumed high quality "background" conditions. However, monitoring data shows atypical water quality results. The station data suggests there is a pollution source causing elevated nutrient loads, low pH conditions and reduced dissolved oxygen. Association observations suggests the pollution plume originates from the old toilet vaults area in the Summit Lake parking area and this pollution plume affects data results from sites 36 and 37.

In June 2012, the Association began a special study effort to document the extent and magnitude of the potential pollution plume. The Association walked the area to identify potential problem areas. The Association concluded that the source area was in the vicinity of the parking lot. The Association originally assumed the problem was related to the past waste disposal practice of using pit privies. There are many wetland or bog ponds that occur between the parking lot and upper Bear Creek. The Association noted that some of these ponds in a drainage fall-line had much more algal productivity than those ponds nearer Summit Lake and well downstream from the parking lot. This suggested that the pollution plume was surfacing in-part in some of the ponds.

E. Coli Special Monitoring

BCWA Fact Sheet 39 E. coli identifies standards for waters in the Bear Creek Watershed and lists those stream segments on the Colorado 303(d) list of impaired waters. An alternative BCWA management program that doesn't require the adoption of a formal regulatory total maximum daily load for E. coli in these listed segments or other

waterbodies in the watershed with suspected bacterial contamination contains the following management strategies and approaches.

Waterway Source Tracking

Fecal coliform & *E. coli* bacteria found in streams in the watershed originate in human, pet, livestock, and wildlife waste. Irrigation, stormwater runoff, snowmelt and flood water, failed on-site wastewater treatment systems (OWTS) leach fields, broken/leaking sewer lines contaminated with fecal matter pose higher risks. *E. coli* does not occur naturally in soil and vegetation but can survive for periods in moist soil or on vegetation. It only enters water from fecal contamination.

- 1. Routine bacterial monitoring at long-term reference sites can be used to detect presence of E. coli over established standards (in lower watershed two-sites below BCR and near Wadsworth on Bear Creek). Routine sites include comprehensive water quality monitoring as defined in the BCWA annual water quality sample plan using established quality assurance protocols.
- 2. Systematic bacterial monitoring when E. coli is detected can be used to trace upstream potential sources of contamination. A targeted monitoring process can identify and isolate likely problem areas. If a problem area is identified, targeted management solutions for that site can be applied through established watershed partners and land-use decision makers.
- 3. Predict potential pathways from land uses (e.g., map pastures, large animal grazing or corral operations, parks and open space, dog parks, locate OWTS by sub-drainages, maps of sewer mains, erosional problem areas and high use human recreational areas
- 4. Establish a water watch program for citizens and businesses that includes education & trained data collection.
- 5. Apply adaptive management to monitoring program. Provide an annual technical memorandum on E. coli management in the watershed.

Stream Flow Data

Manual Site-Specific Measurements

Manual flow measurements are performed at watershed locations through the program period. The Association uses three methods to obtain flow data:

- 1. Manual stream flow is measured using an OTT MF Pro to measure average water velocity in a stream crosssection. Depth and velocity readings are taken at 2-foot increments in the cross-section. Velocity readings are taken at the mid-water column level. Flow is calculated for each sub-section of the cross-section (Q=V x A) and the subsection flows are added to estimate the stream flow. The manual measures have been checked against the flow measurements taken at both USGS gages and the manual flows are generally within 5% of the USGS estimates.
- 2. The Association has an OTT MF Pro flow meter. When the sensor is placed in flowing water, a magnetic field around the sensor creates a voltage proportional to the flow velocity. This voltage amplitude, which represents the rate of water flow around the sensor, is detected by electrodes in the sensor and processed by the sensor microprocessor. The processed signal is digitally transmitted through the sensor cable to the portable meter and the information is shown on the meter display. The system includes a portable meter, sensor with cable and a depth probe. Velocity is calculated in user defined cross-sections that allows for a more accurate estimate of stream or conduit flow than the flow probe. The Mean-section method divides the cross-section into individual flow segments. Pairs of adjacent verticals are the limits of the segments. The two edges of the cross-section are given values of 0 for the velocity and depth. The total flow is the sum of the partial flows of all segments.

USGS, Colorado Department of Water Resources and Urban Drainage Gaging Station Stream Flows

Flow data summarizes the flow gages located on Bear Creek. There are five locations in the watershed that produce flow data and include above Evergreen Lake, above Morrison, Bear Creek Park (partial year), Turkey Creek and below the dam in the outflow trace.

A USGS stream gage (USGS 06710385) maintains a location above Evergreen Lake, near the CDOW fish survey site. The gage location is adjacent to the Denver Mountain Parks golf course (Keys on the Green) parking lot.

The second gaging station is located below the temperature datalogger location ID MORR10, above the town of Morrison, just west of the Highway 8 Bridge over Bear Creek. This station (BCMORCO 06710500) is maintained by the Colorado Division of Water Resources. Weekly stream flow graphs were printed from both stations and filed for record. Monthly average daily flows from gages exported to a spreadsheet for comparison with historical data. Although flow records began at this location in 1899, the most complete data record exists from 1919 through 2016 for the Morrison gage. A 25-year record recorded in annual data reports.

P3 - Watershed Monitoring Programs

The Bear Creek Watershed Association conducts watershed scale monitoring programs. These P3 monitoring programs focus on characterizing water quality of surface waters in the Turkey Creek and Bear Creek drainages of the watershed. The monitoring year is divided into a warm-season period with more intense sampling and a cold-season period, designed to provide minimal winter and spring data. This data report summarizes temperature and water quality monitoring data, sampling results obtained from in-stream locations, and data from five-wastewater treatment plant (WWTP) effluents. The complete Cold-season and Warm-season water quality data set is an electronic data summary report and spreadsheet.

P3 Field Parameter Probe Measurements

Monthly measurements are performed in the morning and early afternoon. Measurements are recorded with an YSI Professional hand-held meter. The meter utilizes a multi-probe sensor, capable of measuring pH, Temperature, Dissolved Oxygen and Specific Conductance simultaneously. Measurements are logged, retained in the on-board computer, and then manually downloaded. Typically, the logged data is manually downloaded by viewing each file and transcribing data onto monthly Logsheets. The data is entered into a spreadsheet. Prior to the program, the meter is calibrated by certified technicians. Prior to each monitoring event, the meter is calibrated for each parameter, using a purchased calibration solution for specific conductance and purchased pH buffers (two-point calibration, 7.0 and 10.0). Fresh batteries installed in the meter at the start of the program and batteries replaced when the observed battery charge reached 50%. Flow measurements are performed coincidentally with monthly sampling and monitoring. A Global Water flow probe Model FP101 is used and values obtained are combined with stream width and depth measurements to calculate estimated streamflow.

P3 Evergreen Lake Monitoring

Evergreen Lake has a temperature data logger string near the dam structure with temperature logger profile at ice-off (April-May) through November 1: ½ m, 1m, 1 ½m, and 2m. Profile data is collected in May to October for Temperature, DO, specific conductance, pH and Turbidity at 0m, -1m, -2m, -3m, and -4m. Chemical analyses include chlorophyll a, Total Nitrogen, and Total Phosphorus. Water is sampled at -1m and +1m in water column. A total depth and Secchi reading are collected. Chlorophyll is sampled at only -1m. Association may collect data consistent with protection of a major drinking water supply system.

P3 Temperature Datalogger Monitoring Locations

Programmable temperature dataloggers measure and record watershed stream and WWTP effluent temperatures every thirty minutes (Table 19). The loggers used in the Program are Onset Computer Corporation brand, Temp Pro v2 (U22) programmable dataloggers. Every other year all model dataloggers are returned to Onset for a NIST (National Institute of Standards and Technology) one-point certification and a 'tune-up'. The one-point certification is performed against calibration standards at 20°C. The 'tune-up' consists of a new battery and quality control testing, assuring the dataloggers meets manufacturer's operating specifications. The Association maintains a fact sheet with temperature monitoring protocols, as included in the Association annual report.

Table 18WWTF Temperature Data

	Continuous effluent	No continuous effluent
	measurements every half-hour,	temperature data
WWTF	Jan 1-Dec 31	expected
Morrison	Х	
Evergreen Metro District	Х	
West Jefferson County Metro District	Х	
Kittredge Sanitation and Water District	Х	
Genesee Water and Sanitation District	Х	
JCS Outdoor Lab		Х
Forest Hills Metro District		Х
Brookforest Inn		Х
Aspen Park Metro District	If surface Discharging	
Tiny Town		Х
Geneva Glen		Х
Conifer Metro District / Conifer Sanitation Association		Х
JCS Conifer High School	Х	

The dataloggers are programmed for measurements every thirty minutes at an office computer equipped with the Onset software. At this frequency, the memory capacity is approximately 905 days for the U22 (Water Temp Pro) series logger. The Association employs newer models with delayed-start capabilities. Logsheets are utilized to record the exact time of deployment and retrieval of all units, so that erroneous measurements (measurements recorded out of water) can be omitted during the data evaluation process.

The U22 series loggers are utilized in all watershed stream locations. These loggers are downloaded to a shuttle device. Occasionally, the download process occurred precisely at the measurement instance and a measurement is lost. There are no watertight cases required for the U22 model loggers. The date and deployment time for all loggers is noted on a Logsheet. After downloading the last logger in the Watershed, the laptop and shuttles are transported to the desktop computer with the Onset software at the EMD Administration office. The logger data is transferred from the laptop and from the shuttles to the desktop. The shuttles are connected to the computer via a download cable, and data on the shuttles are individually downloaded into separate program files.

30-minute datalogger temperature measurements are exported from the Onset Computer software into Excel spreadsheets. Each download of temperature data is treated as a file in the Onset software. Once the Onset file formats is exported and saved as separate Excel files, the Excel spreadsheets for each location is combined into one Excel spreadsheet with multiple worksheets. Therefore, each Excel file contains multiple worksheets, one for each separate download of data, and a summary worksheet. The master dataset spreadsheet contains separate worksheets for each Site in the watershed, displaying all temperature datalogger values and statistical analysis, as well as sampling and monitoring data and statistics.

The date and time recorded on the Launch/Retrieval Logsheet is used to eliminate erroneous temperature measurements prior to data analysis. The majority of these erroneous measurements are eliminated by utilizing the shuttle devices to field-download data. Occasionally, the field download process occurs exactly at the time of a measurement, and an erroneous value is recorded or missed. These are also removed from the raw data prior to analysis. Once in a spreadsheet format, the data is evaluated against the underlying standard Weekly Average Temperature (WAT) criteria, against the underlying standard Daily Maximum Temperature (DM) criteria and against the Maximum Weekly Average Temperature (MWAT) criteria. Percentages of compliance are calculated. Weekly Average Temperatures are determined by calculating the mean temperature of seven consecutive days of data beginning with either April or May or the first day of data collection. Any lack of data collection resulting in a data gap of one day or more, requires that the seven-day period begin anew. Maximum Weekly Average Temperatures are determined by eraluated Weekly Average Temperatures are determined by evaluating the calculated Weekly Average Temperatures.

Daily Maximum values are obtained by calculating the average temperature of a two-hour period beginning with the first temperature recorded, and determining the maximum value from each day. Again, any lack of data collection resulting in a data gap more than two hours, requires that the two-hour calculation period begin anew. In most cases, there are four measurements in a two-hour period.

Limited Seasonal Temperature Datalogger Monitoring Locations

Continuous temperature measurements taken by loggers every half-hour, May 15-Nov 15 (ice dependent)

- Site 58 Bear Creek below the Mt. Evans Wilderness
- Site 25 Vance Creek
- Site 2a Golden Willow on Upper Bear Creek
- Site 3a Above Evergreen Lake at Keys-on-the-Green, CDOW Site
- Site 4 Evergreen Lake, at dam (1/2m, then at 1-meter intervals)

Seasonal Temperature Datalogger Monitoring Locations

Continuous temperature measurements taken by loggers every half-hour, Apr 15-Dec 1 (ice dependent)

- Site 19 North Turkey Creek Flying J Ranch
- Site 18 South Turkey Creek below APMD
- Site 16a Turkey Creek within Bear Creek Park (Lakewood)
- Site 15a Bear Creek Segment 1b at Gaging Station in Bear Creek Park
- Site 40 Bear Creek Reservoir (1/2m-2m)
- Site 45 Below Bear Creek Reservoir
- Site 90 Bear Creek at Wadsworth
- Site 5 Above EMD WWTP, CDOW Downtown Site
- Site 9 O'Fallon Park (west end, CDOW Site)
- Site 12 Lair o' the Bear, CDOW Site
- Site 13a below Idledale (at Idledale at Shady Lane, CDOW Site)
- Site 14a Morrison Park (west end of town, CDOW Site)

P-4 Colorado Division of Parks & Wildlife Fishery Sites, Macroinvertebrates

Weather (local)

A National Weather Service Cooperative Reporting Station Number 052790 is maintained at the EMD WWTP. Daily high and low air temperatures and precipitation are recorded and transmitted monthly to the National Weather Service. Weather data was tabulated and correlated with Bear Creek stream flows (obtained at the USGS gage above Evergreen Lake) for the Program. Weather data collected during the program period was compared to the available historical weather records, obtained at the NWS High Plains Climate Center.

Fishery Surveys

The Colorado Division of Wildlife (CDOPW) has monitored fish populations in the watershed from 1988 through 2017. Prior to 2005, there were five monitoring sites. In 2010, there were ten survey sites (Table 20). In 2011, Bear Track site 38 was included in survey. All of the CDOPW survey sites incorporated in the Association monitoring network. No fish surveys were done in 2012 due to the low flow conditions. No fish surveys were done in 2013 due to flood conditions. Fishery survey completed in 2014-2017. No survey in September 2018 due to low flow concerns.

Stream Segment	CDOPW Fishery Reference Sites
Segment 3	Vance Creek (one-time)
Segment 1a	Keys on the Green, Golden Willow Bridge, Site 58
Segment 1e	Little Bear, Bear Creek Cabins, O'Fallon Park, Lair O' the Bear, Idledale, Morrison Park
Segment 7	Bear Tracks (one-time)

Table 19	Colorado Division of Parks and Wildlife Fish Survey Sites
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These reference sites have coordinated chemistry, biological, physical data collection. This mixed data analysis establishes reference conditions for four stream segments in the watershed. The CDOPW surveys fish populations in September. The survey determines young of the year and adult size classes, species present, total biomass of fish by species and total pounds per acre by species. The Association assists the CDOPW with fish sampling. The CDOPW provides raw and processed data to the Association for the annual data report.

BCWA Macroinvertebrate and Habitat Sampling

See *BCWA ME04 Macroinvertebrate Field Sample Method* for field collection protocols. The reference sites in Table 12 sampled for macroinvertebrates, physical habitat (modified Rapid Bioassessment Protocol) and streambed characterization (modified Wolman Pebble Count). The WQCD's procedure on physical habitat is a visual assessment of the quality of the

instream and riparian habitat that influences the structure and function of the aquatic community in a stream. Parameters are ranked as optimal, suboptimal, marginal, or poor based on a 4-point scale, with 4 being the best possible (optimal) conditions and one representing the worst (poor) conditions.

Macroinvertebrate samples have been historically collected at 7 CDOPW fish survey sites along Bear Creek: Morrison Park, Idledale, Lair o' the Bear Park, O' Fallon Park, Bear Creek Cabins, Main Street Evergreen (across from the Little Bear), above Evergreen Lake upstream of the USGS gaging station, Golden Willow Bridge site 2a and site 58. The WQCD previously assisted with data analyses and interpretation. The sampling, data analyses and interpretation is now an Association function. The macroinvertebrate sampling is done by the Association in August with analyze done by GEI. Sample collection done by the state timed-kick net methodology protocol (Benthic Macroinvertebrate Sampling Protocols, Water Quality Control Division, Standard Operation Procedure, WQCDSOP-001, May 2010).

Table 21 shows the Association sample locations for macroinvertebrates, physical habitat (modified Rapid Bioassessment Protocol) and streambed characterization (modified Wolman Pebble Count). Due to stream bed alterations from the September 2013 flooding event, the Association did new physical habitat and streambed characterization in September 2015; additional surveys completed in 2016-2018.

Primary Target Sites	Secondary Target Sites
Golden Willow Bridge site 2a	Idledale
Dedisse Park near Keys on the Green	Site 58
Little Bear in downtown Evergreen	O' Fallon Park
Bear Creek Cabins	Wadsworth
Lair o' the Bear Park	Site 45
Morrison (gage)	Site 90
BCLP	Turkey Creek

Table 20 BCWA Macroinvertebrate Stations